Docket No.: 15095-02

Amendment to the Specification:

Please replace paragraph [0138] at page 43 with the following amended paragraph:

[0138] Culturing was carried out in a climatic chamber at an air temperature of 25°C and light intensity of 55 micromols micromol s-1m-2 (white light; Philips TL 65W/25 fluorescent tube) and a light/dark change of 16/8 hours. The moss was either modified in liquid culture using Knop medium according to Reski and Abel (1985, Planta 165:354-358) or cultured on Knop solid medium using 1% oxoid agar (Unipath, Basingstoke, England). The protonemas used for RNA and DNA isolation were cultured in aerated liquid cultures. The protonemas were comminuted every 9 days and transferred to fresh culture medium.

Please replace paragraph [0168] at page 59 with the following amended paragraph:

[0168] T1 seeds were sterilized according to standard protocols (Xiong et al. 1999, Plant Molecular Biology Reporter 17: 159-170). Seeds were plated on ½ Murashige and Skoog media (MS) (Sigma-Aldrich) pH 5.7 with KOH, 0.6% agar and supplemented with 1% sucrose, 0.5 g/L 2-[N-Morpholino]ethansulfonic acid (MES) (Sigma-Aldrich), 50 μg/ml kanamycin (Sigma-Aldrich), 500 μg/ml carbenicillan (Sigma-Aldrich) and 2 μg/ml benomyl (Sigma-Aldrich). Seeds on plates were vernalized for four days at 4°C. The seeds were germinated in a climatic chamber at an air temperature of 22°C and light intensity of 40 micromols—1 m2 micromol s⁻¹ m⁻² (white light; Philips TL 65W/25 fluorescent tube) and 16 hours light and 8 hours dark day length cycle. Transformed seedlings were selected after 14 days and transferred to ½ MS media pH 5.7 with KOH 0.6% agar plates supplemented with 0.6% agar, 1% sucrose, 0.5 g/L MES (Sigma-Aldrich), and 2 μg/ml benomyl (Sigma-Aldrich) and allowed to recover for five-seven days.

Please replace paragraph [0169] at page 60 with the following amended paragraph:

[0169] T1 seedlings were transferred to dry, sterile filter paper in a petri dish and allowed to desiccate for two hours at 80% RH (relative humidity) in a Percival Growth CU3615, $\frac{1}{1000} = \frac{1}{1000} = \frac{1}{$

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Please replace paragraph [0172] at page 61 with the following amended paragraph:

[0172] Seedlings were moved to petri dishes containing $\frac{1}{2}$ MS 0.6% agar supplemented with 2% sucrose and 2 µg/ml benomyl. After four days, the seedlings were incubated at 4°C for 1 hour and then covered with shaved ice. The seedlings were then placed in an Environmental Specialist ES2000 Environmental Chamber and incubated for 3.5 hours beginning at -1.0° C decreasing -1.0° C hour 1° C per hour. The seedlings were then incubated at -5.0° C for 24 hours and then allowed to thaw at 5°C for 12 hours. The water was poured off and the seedlings were scored after 5 days.

Please replace paragraphs [0188]-[0191] at pages 65-66 with the following amended paragraphs: [0188] The constructs pBPSLVM162, pBPSJYW004, pBPSJYW001, pBPSLVM180 and pBPSJYW006 were are used to transform soybean as described below.

[0189] Seeds of soybean were are surface sterilized with 70% ethanol for 4 minutes at room temperature with continuous shaking, followed by 20% (v/v) Clorox supplemented with 0.05% (v/v) Tween for 20 minutes with continuous shaking. Then, the seeds were are rinsed 4 times with distilled water and placed on moistened sterile filter paper in a Petri dish at room temperature for 6 to 39 hours. The seed coats were are peeled off, and cotyledons are detached from the embryo axis. The embryo axis was is examined to make sure that the meristematic region is not damaged. The excised embryo axes were are collected in a half-open sterile Petri dish and air-dried to a moisture content less than 20% (fresh weight) in a sealed Petri dish until further use.

[0190] Agrobacterium tumefaciens culture was is prepared from a single colony in LB solid medium plus appropriate antibiotics (e.g. 100 mg/l streptomycin, 50 mg/l kanamycin) followed by growth of the single colony in liquid LB medium to an optical density at 600 nm of 0.8. Then, the bacteria bacterial culture was is pelleted at 7000 rpm for 7 minutes at room temperature, and resuspended in MS (Murashige and Skoog, 1962) medium supplemented with 100 µM acetosyringone. Bacterial cultures were are incubated in this pre-induction medium for 2 hours at room temperature before use. The axis axes of soybean zygotic seed embryos at approximately 15% moisture content were are imbibed for 2 hours at room temperature with the pre-induced Agrobacterium suspension culture. The embryos are removed from the imbibition culture and were are transferred to Petri dishes containing solid MS medium supplemented with

2% sucrose and incubated for 2 days, in the dark at room temperature. Alternatively, the embryos were are placed on top of moistened (liquid MS medium) sterile filter paper in a Petri dish and incubated under the same conditions described above. After this period, the embryos were are transferred to either solid or liquid MS medium supplemented with 500 mg/L carbenicillin or 300mg/L cefotaxime to kill the agrobacteria. The liquid medium was is used to moisten the sterile filter paper. The embryos were are incubated during for 4 weeks at 25°C, under 150 μmol m⁻²sec⁻¹ and 12 hours photoperiod. Once the seedlings produced roots, they were are transferred to sterile metromix soil. The medium of the *in vitro* plants was is washed off before transferring the plants are transferred to soil. The plants were are kept under a plastic cover for 1 week to favor the acclimatization process. Then the plants were are transferred to a growth room where they were are incubated at 25°C, under 150 μmol m⁻²sec⁻¹ light intensity and 12 hours photoperiod for about 80 days.

[0191] The transgenic plants were are then screened for their improved drought, salt and/or cold tolerance according to the screening method described in Example 7 demonstrating to demonstrate that transgene expression confers stress tolerance.

Please replace paragraph [0192] at page 66 with the following amended paragraph:

[0192] The constructs pBPSLVM162, pBPSJYW004, pBPSJYW001, pBPSLVM180 and pBPSJYW006 were are used to transform rapseed rapeseed/canola as described below.

Please replace paragraph [0195] at page 67 with the following amended paragraph:
[0195] The constructs pBPSLVM162, pBPSJYW004, pBPSJYW001, pBPSLVM180 and pBPSJYW006 were are used to transform corn as described below.

Please replace paragraph [0197] at page 67 with the following amended paragraph: [0197] The constructs pBPSLVM162, pBPSJYW004, pBPSJYW001, pBPSLVM180 and pBPSJYW006 were are used to transform wheat as described below.